

Physiological fluorescence lifetime imaging microscopy improves Förster resonance energy transfer detection in living cells

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Abstract. Accurate, unambiguous detection of molecular interactions in living cells via measurements of Förster (or fluorescence) resonance energy transfer (FRET) events is experimentally challenging. We develop and apply a physiological fluorescence lifetime imaging microscopy (physiological FLIM) system to significantly improve FRET detection in living cells. Multiple positive and negative cellular controls are implemented to validate the experimental method developed. FLIM measurement techniques were found to remove fluorescence intensity-based artifacts, resulting in a seven-fold improvement in fluorescence measurement precision. The addition of cellular environmental controls, including both temperature and CO₂ stabilization, for physiological FLIM eliminates non-specific FRET in the live-cell system studied. Overall, only physiological FLIM results in statistically significant results that clearly indicated the presence of specific molecular interactions in the live-cell system. This approach can be applied generally to improve the accuracy and precision of FRET measurements in living cells. © 2009 Society of Photo-Optical Instrumentation Engineers. [DOI: 10.1117/1.3257254]

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Förster (or fluorescence) resonance energy transfer (FRET) can be used as an *in vivo* nanoscale ruler to detect the distance between fluorophores or interacting proteins with attached fluorophores.¹ In intensity-based FRET detection, sophisticated calibration and correction procedures are usually required to cope with intensity variations, spectral bleedthrough,² back-bleedthrough,³ and photobleaching.⁴ In addition, nonspecific FRET, arising from random collisions of donor and acceptor and/or other unexpected associations of donor and acceptor, affects FRET accuracy, and although it can be detected, it may not be easily removed.⁵

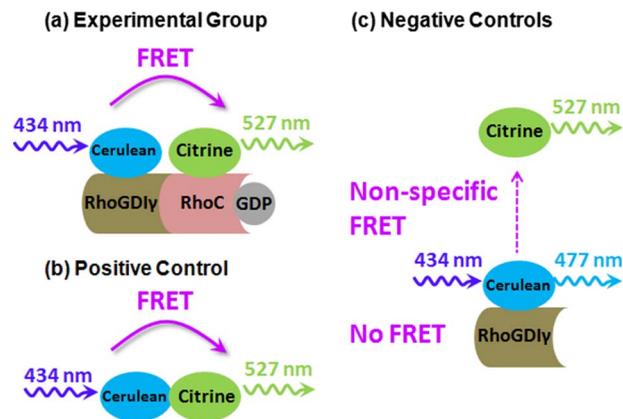


Fig. 1 Schematic depiction of FRET studies on RhoC and RhoGDI_γ. (a) The experimental cellular group cerulean-RhoGDI_γ+citrine-RhoC (with FRET). (b) The positive control cellular group cerulean-citrine (with FRET). (c) Two negative control cellular groups: cerulean-RhoGDI_γ alone (no FRET) and cerulean-RhoGDI_γ+citrine (nonspecific FRET). Fluorophore excitation and emission maxima are labeled.

Fluorescence lifetime imaging microscopy (FLIM) retrieves fluorophore lifetime data from fluorescence emission, and can be used to detect FRET with some advantages over intensity-based FRET.⁶ Interest in FLIM-FRET is growing, especially as commercial FLIM modules become available for confocal and multiphoton microscopy. However, accurate, unambiguous detection of molecular interactions in living cells remains experimentally challenging due to the presence of nonspecific FRET.

In this study, we develop and apply a physiological FLIM system with experimental controls for CO₂ and temperature stabilization to create a physiological environment for live-cell FLIM studies. Our purpose was to investigate how physiological FLIM affects FRET detection in living cells. To accomplish this, we studied a live-cell system involving Ras Homology Protein C (RhoC) inactive-form interactions. RhoC has been identified as a specific oncogene marker of aggressive breast cancers.⁷ When active, it is anchored on the cell membrane, interacting with effectors. When inactive, it is bound to its inhibitor RhoGDI_γ in the cytoplasm.⁸ Interactions between RhoGDI_γ and RhoC caused FRET to occur from the donor (cerulean, attached to RhoGDI_γ) to the acceptor (citrine, attached to RhoC) in our experimental cellular group, as in Fig. 1(a). In the positive control cellular group, Fig. 1(b), cerulean was linked to citrine, and therefore FRET must occur. Two negative control cellular groups [Fig. 1(c)] were implemented: cerulean-RhoGDI_γ alone, which had no FRET occurring, and cerulean-RhoGDI_γ+citrine, which indicated nonspecific FRET. Living CV1 cells (monkey kidney epithelial cells) were either single- or double-transfected with the plasmids encoding corresponding fusion proteins using GeneJammer transfection reagent (Stratagene, La Jolla, California).⁸

The time-domain FLIM system employed a tunable picosecond laser for excitation, an ultrafast gated, intensified charge-coupled device (CCD) camera for detection, and achieved 50-ps lifetime discrimination.⁹ In this study, cer-

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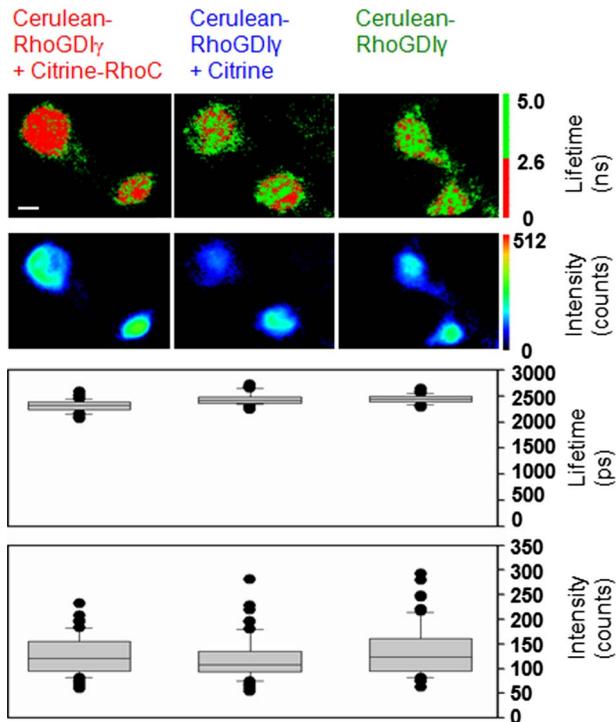


Fig. 2 Comparison of donor fluorescence lifetime vs intensity measurements for live-cell FRET. Representative images (top) and box plots of extracted values after pixel data were averaged per cell (bottom) for the FRET experimental cellular group (left) and the two negative control cellular groups (middle and right). The donor lifetime values of the experimental cellular group were significantly smaller than the two negative controls (p -value $\leq 4.0 \times 10^{-10}$), indicating FRET. This distinction was not discernable from intensity measurements, due to large inter- and intracellular variations (p -value > 0.18). Scale bar = 15 μm .

ulean fluorescence was excited at $\lambda_{\text{ex}} = 436 \pm 10$ nm using the laser dye Coumarin 440 and collected at $\lambda_{\text{em}} = 480 \pm 20$ nm. Because cerulean lifetime may decrease under prolonged irradiation,¹⁰ we eliminated this potential effect by exciting all cellular groups similarly and with short excitation times (< 10 s for each cell). In retrieving lifetime values, a four-gate protocol with a linearized least-square fitting method was used.⁸ The gate width was 6.4 ns and the time interval between two consecutive gates was 1.6 ns. Temperature control at 37 ± 0.2 °C was achieved by using Delta T dishes along with a plate heater and an objective heater (Bioptechs, Incorporated, Butler, Pennsylvania) for all studies reported here. The flow of CO₂ into the Delta T dishes was controlled by a peristaltic pump (Model P720, Instech Laboratories, Incorporated, Plymouth Meeting, Pennsylvania) connected to the Delta T perfused heated lid (Bioptechs, Incorporated) enclosing the dishes. The heated lid had two ports, which could be used for perfusion. When the flow rate was approximately 11 ml/hr, the pH value in the cell culture media became stable at ~ 7.4 in 10 min and remained stable for at least 90 min. Physiological data were acquired under these conditions. The achieved stable pH value could be adjusted by increasing or decreasing the flow rate, while the absence of CO₂ flow caused the pH value to reach ~ 8.4 in 10 min and to increase continuously for at least 90 min. To statistically compare the mean values of FRET donor lifetimes or intensities

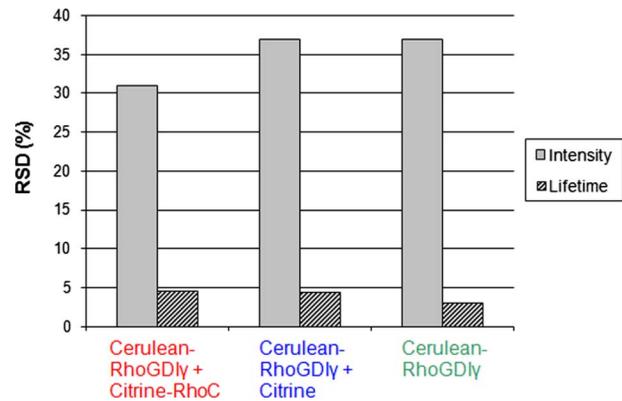


Fig. 3 Precision of donor fluorescence intensity (gray bars) vs lifetime (slash-shaded bars) measurements as quantified by relative standard deviation (RSD) values for the FRET experimental cellular group (left) and the two negative control cellular groups (middle and right). Precision improved roughly seven-fold when fluorescence lifetime data were used rather than fluorescence intensity data. RSD = standard deviation/mean. The number of cells measured in each group is labeled at the top.

among cellular groups, pixel data were averaged per cell and two-tailed student's t -tests were used. We employed a conventional significance level of 0.1, even though we always obtained much smaller p -values for stronger confirmation of statistical significance.

Figure 2 compares donor fluorescence lifetime versus intensity measurements for live-cell FRET. Both temperature and CO₂ controls were implemented. The donor lifetime values of the experimental cellular group were significantly smaller than the two negative controls (p -value $\leq 4.0 \times 10^{-10}$), indicating FRET. The positive control cellular group (data not shown) also exhibited a shorter lifetime than the two negative controls, as expected. Intensity images, however, exhibited many more inter- and intracellular variations, thereby obscuring the detection of any possible FRET (both specific and nonspecific) occurring in this system. Figure 3 compares the precision of donor fluorescence intensity versus lifetime measurements, as quantified by relative standard deviation (RSD) values obtained from the FRET data in Fig. 2, and reveals the multifold improvement in precision when fluorescence lifetime data were used rather than fluorescence intensity data.

Table 1 presents p -values from t -test comparisons among cellular groups, and demonstrates that only physiological FLIM led to statistically significant FRET results without being influenced by nonspecific FRET. The temperature was fixed at 37 °C, while CO₂ control was varied and both intensity and FLIM data were acquired. With physiological FLIM, the experimental group versus either negative control comparison had very small p -values ($\leq 4.0 \times 10^{-10}$), which suggested molecular interactions were detected by FRET, while the comparison of the two negative controls gave a large p -value (0.93), which suggested that no nonspecific FRET between cerulean and citrine occurred. Without physiological conditions, significant nonspecific FRET did occur, as indicated by the statistically significant p -value from the comparison of the two negative controls for both FLIM and intensity data (9.43×10^{-5} and 4.50×10^{-3} , respectively), and thus affected the accuracy of any FRET detected.

Table 1 The p -values from t-test comparisons among cellular groups. The temperature was fixed at 37 °C, while CO₂ control was either implemented (left column under p -values) or not implemented (right column under p -values). Only physiological FLIM led to statistically significant results without nonspecific FRET.

t-test comparison groups			p-values	
Experimental		Negative control	Physiological FLIM physiological intensity	FLIM intensity
Cerulean-RhoGDI γ + citrine-RhoC	vs	Cerulean-RhoGDI γ +citrine	4.00×10^{-10}	3.07×10^{-5}
			0.44	8.85×10^{-4}
Cerulean-RhoGDI γ + citrine-RhoC	vs	Cerulean-RhoGDI γ	$<1.00 \times 10^{-10}$	$<1.00 \times 10^{-10}$
			0.19	0.89
		Cerulean-RhoGDI γ +citrine	0.93	9.43×10^{-5}
			Cerulean-RhoGDI γ	5.67×10^{-2}

Due to their high variability, intensity-based measurements also led to inconclusive results regarding the detection of molecular interactions. Physiological intensity measurements showed no statistically significant differences between the experimental cellular group and either negative control. The large p -values obtained (0.44 and 0.19) reflected the large distributions of intensities of the FRET donor (Fig. 2) that masked the presence of molecular interactions. We attribute the slightly significant p -value for the negative control comparison (5.67×10^{-2}) to these large intensity variations.

Since the fluorescence intensity-based data presented here were not acquired and processed with sophisticated FRET calibration and correction procedures,^{2,3} the comparisons of intensity and lifetime measurements should not be regarded as comparisons of intensity-based FRET detection and lifetime-based FRET detection. Rather, the key point is that lifetime-based FRET detection is relatively simple to implement and accomplish, without complicated corrections, while still offering statistically significant evidence for FRET.

Nonspecific FRET is thought to arise from random collisions and/or other unexpected associations of donor and acceptor.⁵ Here, CO₂ control stabilized the pH value of the cell culture media surrounding the cells at physiological values, and was found to eliminate nonspecific FRET in the live-cell FRET-FLIM experiments. Significant nonspecific FRET was observed in experiments conducted without CO₂ control, when media pH values rose to unphysiological levels. It has been reported that nonspecific FRET was absent in a live-cell system when physiological conditions were employed,¹¹ and we further demonstrated that these physiological conditions could be responsible for the elimination of nonspecific FRET in live-cell systems. We also observed somewhat higher variability in donor lifetime distributions for all cellular groups when CO₂ was uncontrolled (average RSD=4.76%) versus when CO₂ control was implemented (average RSD=4.11%). Given the potential influence of pH on protein stability, enzyme and ion channel activity, molecular conformation, and many other cellular and molecular processes, careful regulation of the cellular environment through physiological FLIM offers a valuable tool for FRET studies in living cells.

Implementing physiological FLIM significantly improved FRET detection, clearly and unambiguously indicating the presence of specific molecular interactions between RhoGDI γ and RhoC in the live CV1 cells studied. The approach described here is generally applicable and should improve FRET detection in a variety of live-cell systems.

Acknowledgments

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