Biomedical Optics

SPIEDigitalLibrary.org/jbo

Evaluation through *in vivo* reflectance confocal microscopy of the cutaneous neurogenic inflammatory reaction induced by capsaicin in human subjects

Constantin Căruntu Daniel Boda



Evaluation through *in vivo* reflectance confocal microscopy of the cutaneous neurogenic inflammatory reaction induced by capsaicin in human subjects

Constantin Căruntua and Daniel Bodab

^aCarol Davila University of Medicine and Pharmacy, Center for Excellence in Neuroscience, 8 Eroii Sanitari Blvd., 050474 Bucharest, Romania ^bCarol Davila University of Medicine and Pharmacy, Dermato-oncology Research Laboratory, 22-24 Gr. Manolescu, 0111234, Sector 1, Bucharest, Romania

Abstract. We perform an *in vivo* analysis of the effects of capsaicin on cutaneous microvascularization. A total of 29 healthy subjects are administered a solution of capsaicin (CAP group) or a vehicle solution (nonCAP group) on the dorsal side of the nondominant hand. The evaluation is performed using *in vivo* reflectance confocal microscopy (RCM). Ten minutes after administration, the area of the section, the perimeter, and the Feret's diameter of the capillaries in the dermal papillae become significantly larger in the CAP group as against the nonCAP group, and this difference is maintained until the conclusion of the experiment. *In vivo* RCM allows the investigation of cutaneous vascular reactions induced by capsaicin. As such, this method may constitute an useful technique both for research and clinical practice. © 2012 Society of Photo-Optical Instrumentation Engineers (SPIE). [DOI: 10.1117/1.]BO.17.8 .085003]

Keywords: neurogenic inflammation; capsaicin; in vivo reflectance confocal microscopy; microvascularization; skin.

Paper 12223 received Apr. 9, 2012; revised manuscript received Jun. 16, 2012; accepted for publication Jul. 17, 2012; published online Aug. 13, 2012.

1 Introduction

Neurogenic inflammation is the inflammatory process whose inductor mechanism is the activation of the nerve fibers. This type of reaction may have a contributing role in the development and aggravation of psoriasis, ^{1–3} atopic dermatitis, ^{3–6} urticaria, ¹ and perioral dermatitis. ⁷ Neurogenic inflammation has also been linked to the formation of keloids and hypertrophic scars ⁸ and to the early stages of rosacea. ⁹ Neurogenic-mediated inflammation increasingly appears to be a factor in the onset and evolution of various skin diseases, ^{4,5,10–12} and the study of interactions between the nervous system and the skin is of great interest for the scientific world.

Neurogenic cutaneous inflammation induced by local administration of capsaicin, the hot ingredient of red pepper, is one of the most used study models. In the skin, capsaicin causes a painful burning-like sensation and a erythematous-edematous inflammatory reaction. These effects are primarily induced by its direct action on the transient receptor potential vanilloid subtype 1 (TRPV1) channel found in the nerve endings of type C fibers. As a consequence of capsaicin action on nerve fibers, proinflammatory neuropeptides are released, especially substance P (SP) and calcitonin gene-related peptide (CGRP), shift initiate an inflammatory process associated with cutaneous vasodilation, an increase of vascular permeability, plasmatic extravasation, and edema.

Capsaicin-induced neurogenic inflammation, by either topical administration^{20–25} or local injection, ^{13,26,27} has been used in many research projects on human subjects. Topical

administration of capsaicin on skin determines a long-term response, as shown in a study where maximum increase in cutaneous blood flow was achieved after 30 to 45 min from application and was maintained even after 1 h.²⁸ It is an easy, noninvasive method with minimal discomfort for the study subjects. Administration of capsaicin by injection provides good control on the dose of active substance, but it is invasive and creates discomfort for the subjects.

In previous studies, methods like infrared thermography,²⁷ laser Doppler flowmetry,^{20–22,26} or marking and measurement of the erythema area on skin with planimetry^{13,27} were used to evaluate capsaicin's effects on cutaneous microvascularization.²⁹ However, none of these methods allowed a histomorphological evaluation of capsaicin's effects on cutaneous microcirculation, and there is little information in scientific literature regarding this aspect.

Reflectance confocal microscopy (RCM) allows a non invasive investigation of cutaneous tissue at a depth of up to 250 μ m with a resolution similar to histological examination. This method allows *in vivo* study of microscopic structures, including some cellular details, in skin layers and real-time observation of different micromorphological parameters. ^{30–32} It is considered an excellent method to evaluate the capillary vessels situated in the dermoepidermal junction, allowing the observation of blood cell dynamics in capillaries and providing the means to analyze the morphology of capillary ansae in dermal papillae. ^{33–36}

This study analyzes the effect of capsaicin on cutaneous microvascularization in human subjects using *in vivo* RCM.

Address all correspondence to: Daniel Boda, Carol Davila University of Medicine and Pharmacy, Dermato-oncology Research Laboratory, 22-24 Gr. Manolescu, 0111234, sector 1, Bucharest, Romania. Tel: +40-757079117; Fax: +40-212221310; E-mail: daniel.boda@umf.ro.

0091-3286/2012/\$25.00 © 2012 SPIE

2 Materials and Methods

2.1 Subjects

A total of 29 healthy subjects (M = 14, F = 15), ages 18 to 35 (average age: 22.62 years), were enrolled in the study. All subjects graduated or were enrolled in a higher education program, and they participated in the study on a voluntary basis. All subjects signed a written consent after having been fully informed about the study, the confidentiality criteria, the rights, and the ethics criteria.

Those with cardiovascular or respiratory diseases, autoimmune diseases, neoplasia, organ transplant, psychiatric disorders, infectious diseases during the previous month, or allergic reactions to the substances used in the study or their derivatives were excluded from the study, as well as those taking medication that might influence the physiological parameters under study. Pregnant or breast feeding women were also excluded from the study.

Study participants were asked to avoid psychoactive substances, alcohol, coffee, tea, energy drinks, beverages containing caffeine, smoking, taking drugs, and strenuous physical effort 24 h before being tested.

The subjects included in the study were randomly divided into two groups. A test group with 15 subjects (M = 8, F = 7) was administered locally capsaicin dissolved in a vehicle solution (capsaicin treated group—CAP), and a control group with 14 subjects (M = 6, F = 8) was administered locally only the vehicle (noncapsaicin treated group – nonCAP).

2.2 Performance of the Experiment

The study was conducted in the Dermato-oncology Research Laboratory of Carol Davila Medicine and Pharmacy University, Bucharest, after the approval of the local Ethics Committee.

The experiments were performed in the afternoon (12 p.m. to 6 p.m.) at a room temperature of $22 \pm 1^{\circ}$ C and a humidity of $50 \pm 5\%$. They lasted approximately 2 h for each subject. Following their arrival at the laboratory, the subjects were seated comfortably and given a period of 45 min to adjust to the local conditions. During that time, they filled in the forms of inclusion in the study, and they watched a slide show on the monitor.

At the end of the adjustment period, the region to be investigated was delimited on the dorsal side of the nondominant hand. Any previous injuries were avoided, as were any maneuvers that might cause an inflammatory reaction at the investigated region. Cutaneous microvascularization was evaluated using *in vivo* RCM. Two sessions of measurements were performed for every subject. Initial evaluation allowed the identification of a baseline level for the parameters under study. The second measurement session was performed after the topical administration of the capsaicin solution (for subjects in the CAP group) or the vehicle administration (for subjects in non-CAP group) on the investigated area. Cutaneous microvascularization changes in the investigated area were quantified at 0, 10, 25, and 40 min after the beginning of administration.

2.3 Administration of Substances on the Investigated Region

Capsaicin (M-2028; Sigma Chemical Co, St Louis, MO) was dissolved at a concentration of 1% in the immersion oil (Crodamol STS oil; Croda Inc., Edison, NJ) used in a standard manner for image acquisition with *in vivo* RCM. A volume of 7.5 μ L

capsaicin solution in immersion oil was administered with a pipette (Biohit Proline Single-Channel Pipettor, Variable Volume, 0.5 to 10 μ L) on the investigated area in the CAP group. The investigated region was isolated from the adjacent skin with a plastic adhesive disc attached to a metallic ring, which was used to fix the image acquisition device. A similar method of administration was used for subjects in the nonCAP group, but the immersion oil did not contain capsaicin.

2.4 Image Acquisition by RCM

We used the VivaScope® 1500 (Lucid Inc, Rochester, NY) to acquire the images taken by RCM. The wavelength of the laser source was 830 nm, which enabled the visualization of structural details in cutaneous microvascularization at the dermoepidermal junction. The magnetic objective of the microscope was placed in contact with the metallic ring fixed to the skin, without applying any pressure.

We acquired sets of images from the center of the investigated region with an area of 4×4 mm, situated in the dermoepidermal junction, for every evaluation stage. Although previous studies did not show an influence of image acquisition depth on morphological parameters of papillary capillaries, ³⁵ in our study, images were acquired at a similar depth for every subject throughout the experiment.

2.5 Cutaneous Microvascularization Evaluation

Images taken by RCM were analyzed using the Java-based image processing and analysis program ImageJ 1.45 (http://rsbweb.nih.gov/ij/), which can be downloaded and used free of charge for scientific research.

For every image, we identified the papillae with visible capillary lumina. After marking the capillary lumina margins, the program allowed automatic calculation of micromorfological parameters. The capillary section area, perimeter, and Feret's diameter (maximum caliper) of capillaries from the dermal papillae were evaluated (see Fig. 1) for a minimum of 14 dermal papillae per subject.

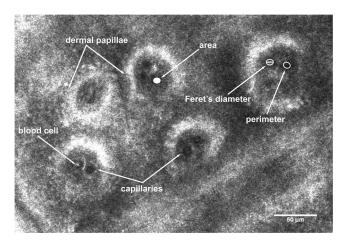


Fig. 1 With *in vivo* RCM images, dermal papillae apear as dark roundoval areas corresponding to dermal tissue, surrounded by bright rings representing cells from the basal layer of the epidermis. Capillary ansae in the structure of dermal papillae appear in transversal section as very dark discs, in which some bright elements can sometimes be noticed representing blood cells. High resolution of the images acquired with RCM enables the micromorphological evaluation of papillary vascular ansae using as parameters the area of section, perimeter, and Feret's diameter of the capillaries.

The confocal microscopy images were evaluated in a blinded manner.

2.6 Statistical Analysis

Values of the investigated parameters were calculated for every subject and for every evaluation stage. Values obtained during the time intervals of 0, 10, 25, and 40 min from administration were expressed as a percentage against the baseline value.

The program SPSS 12.0 (SPSS, Chicago, IL) was used for statistical analysis of data.

Differences between the sexes, and testing of equivalence between the two groups regarding the mean baseline values for each of investigated parameters, were analyzed with one-way between groups analysis of variance (ANOVA). One-way repeated measures ANOVA, followed by Tukey post hoc tests, were used to quantify differences within the two groups for values calculated at every time interval against the baseline for each investigated parameter. When statistically significant differences were revealed, evaluation of correlations existing between these changes was performed, and correlation with the mean baseline values was tested using the Pearson Correlation test.

One-way between groups ANOVA was used for comparative analysis between the two groups for the changes calculated against the baseline value for every time interval.

The results were presented as an average \pm SD. A *P* value <0.05 was considered significant.

3 Results

3.1 Mean Baseline Values

We performed an overall analysis of mean baseline values for each investigated parameter to evaluate possible differences between female and male subjects, and no statistically significant differences were found (all P values > 0.81, ANOVA), as shown in Table 1.

Likewise, possible differences in mean baseline values for the investigated parameters between the two groups were analyzed, and no statistically significant differences were revealed (all P values > 0.13, ANOVA), as shown in Table 2.

3.2 Changes of Investigated Parameters After Capsaicin or Vehicle Solution Application

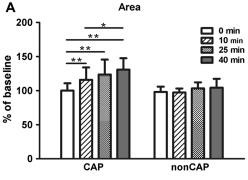
We analyzed the values of investigated parameters at 0, 10, 25, and 40 min from administration, searching possible

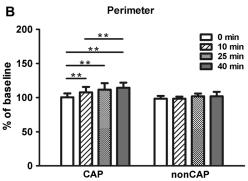
Table 1 Comparative analysis between female and male subjects of the mean baseline values of the investigated parameters.

| | Male subjects | | Female | C· ·(· | |
|---------------------|---------------|-----------------------|---------------|-----------------------|-------------------------------------|
| Parameter | Mean value | Standard deviation | Mean value | Standard deviation | Significance level (P, ANOVA) |
| Area | 44.151 | 19.203 | 42.615 | 14.787 | 0.81041 |
| Perimeter | 22.561 | 5.200 | 22.338 | 4.056 | 0.89810 |
| Feret's diameter | 8.227 | 1.793 | 8.149 | 1.356 | 0.89487 |

Table 2 Comparative analysis between the two groups of the mean baseline values of the investigated parameters.

| | nonCAP | | CAP | | C::[: | |
|---------------------|---------------|-----------------------|--------|-----------------------|-------------------------------------|--|
| Parameter | Mean value | Standard deviation | | Standard deviation | Significance level (P, ANOVA) | |
| Area | 38.522 | 16.514 | 47.869 | 16.245 | 0.13614 | |
| Perimeter | 21.184 | 4.380 | 23.623 | 4.549 | 0.15350 | |
| Feret's diameter | 7.854 | 1.499 | 8.498 | 1.590 | 0.27253 | |





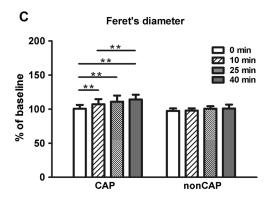


Fig. 2 Analysis performed within each group regarding the changes of parameters at 0, 10, 25, and 40 min. (a) area of the section, (b) perimeter, and (c) Feret's diameter of capillaries did not change significantly in the nonCAP group, whereas in the CAP group, a significant elevation was registered 10 min after the application of capsaicin, followed by a slower increasing trend after 25 and 40 min. Error bars represent the standard deviation. *P < 0.05, **P < 0.01, Tukey post hoc test.

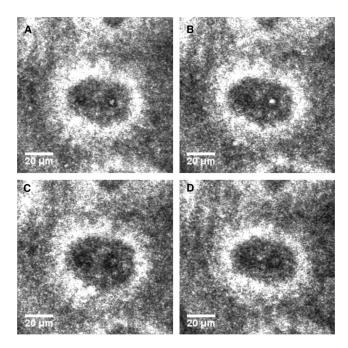


Fig. 3 Images of RCM of the same dermal papilla acquired (a) 0 min, (b) 10 min, (c) 25 min, and (d) 40 min after the application of capsaicin. Dilation of the lumina of papillary capillaries can be noticed after 10 min. This effect becomes stronger 25 and 40 min after the application of the active substance.

differences between sexes, and similar values were obtained, without revealing statistically significant differences for any parameter.

Subsequently, we evaluated within each group the changes of parameters associated to every time interval, and we drew an analysis of correlations between them. The nonCAP group had relatively constant values for capillary section area, perimeter, and Feret's diameter at 0, 10, 25, and 40 min, without revealing any statistically significant differences (see Fig. 2).

By contrast, the CAP group had a significant increase of these parameters 10 min after capsaicin application, followed by a slower increase trend after 25 min and 40 min, as shown in Figs. 2(a) to 2(c) and 3. This increase was not influenced by baseline values of the above mentioned parameters, since no correlations were revealed between baseline values and changes found at the investigated time intervals (Pearson coefficient between -0.09560 and 0.25327; all P values > 0.36241). However, in this group, strong positive correlations were found between values calculated at time intervals of 10, 25, and 40 min after capsaicin application for all parameters (Pearson coefficient between 0.70501 and 0.83267; all *P* values < 0.00334). Subjects with a more intense reaction to capsaicin administration at 10 min maintained a similar response trend at 25 and 40 min after active substance application. These changes in dermal vascular circulation can be easily monitored in real-time examination using RCM (see Videos 1 to 4).

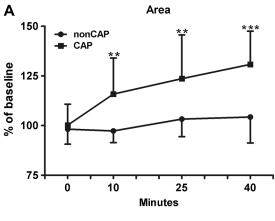
For every time interval, a comparative analysis of values calculated for capillary section area, perimeter, and Feret's diameter was conducted between the two groups (see Table 3 and Fig. 4). Micromorphological evaluation showed a rapid vaso-dilation induced by capsaicin. As early as 10 min after active substance application, the capillary section area, perimeter, and Feret's diameter became significantly larger in the CAP group, and this difference was maintained until the experiment's conclusion.

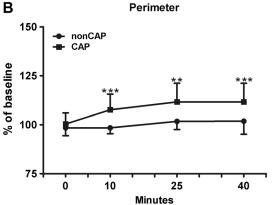
4 Discussions and Conclusions

Our study presents for the first time a micromorphological in vivo evaluation of capsaicin's effects on cutaneous

Table 3 Comparative analysis between the two groups of the values at every moment of time of the investigated parameters expressed as a percentage against their baseline value.

| | | nonCAP | | CAP | | |
|------------------|------------------|---------------|-----------------------|---------------|-----------------------|-------------------------------|
| Time interval | Parameter | Mean value | Standard deviation | Mean value | Standard deviation | Significance level (P, ANOVA) |
| O min | Area | 98.196 | 7.575 | 100.206 | 10.574 | 0.56360 |
| | Perimeter | 98.448 | 4.016 | 100.380 | 5.656 | 0.30132 |
| | Feret's diameter | 97.426 | 3.697 | 100.589 | 5.566 | 0.08488 |
| 10 min | Area | 97.303 | 5.897 | 115.872 | 18.124 | 0.0011 |
| | Perimeter | 98.454 | 2.979 | 107.597 | 7.959 | 0.0004 |
| | Feret's diameter | 97.990 | 3.189 | 107.071 | 7.521 | 0.00028 |
| 25 min | Area | 103.277 | 8.903 | 123.610 | 21.991 | 0.00334 |
| | Perimeter | 101.760 | 4.177 | 111.592 | 9.576 | 0.00148 |
| | Feret's diameter | 100.570 | 3.775 | 111.005 | 8.831 | 0.00035 |
| 40 min | Area | 104.286 | 13.034 | 130.799 | 16.714 | 0.000061 |
| | Perimeter | 101.844 | 6.644 | 114.520 | 7.328 | 0.000043 |
| | Feret's diameter | 100.879 | 5.796 | 114.261 | 6.876 | 0.00005 |





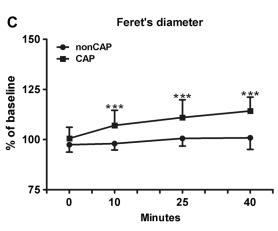
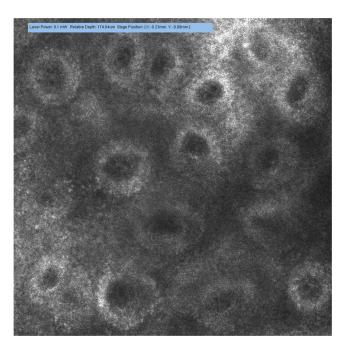


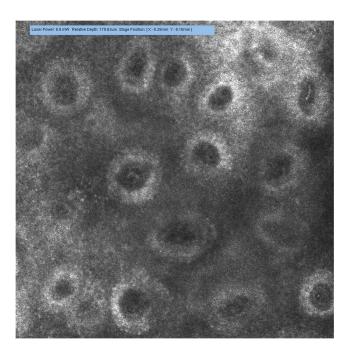
Fig. 4 Comparative analysis between the two groups of values calculated for the investigated parameters for every time interval. (a) Area of section, (b) perimeter, and (c) Feret's diameter of capillaries were significantly higher in the CAP group as against the the nonCAP group as early as after 10 min, these differences were mantained after 25 and 40 min. Error bars represent the standard deviation. **P < 0.01, and ***P < 0.001, one way ANOVA test.

microcirculation. This study enabled us to report the changes induced by capsaicin at different time intervals compared to baseline values for vascular parameters, and it facilitated the drawing of a time curve of vascular reaction induced by capsaicin.

In vivo RCM allows the examination of cutaneous structures to a depth of approximately 250 μ m and investigation of vascular reactions in superficial layers of the skin. There are studies suggesting that, in human subjects, neurogenic inflammatory reaction is not homogeneous in superficial and deep dermis,

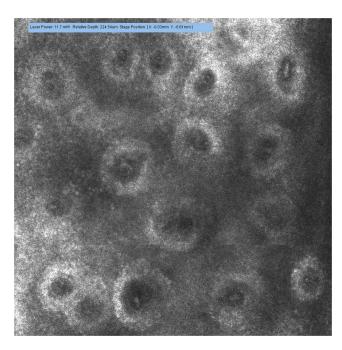


Video 1 Papillary dermis vascular circulation at 0 min from capsaicin application (MPEG, 4.75 MB) [URI: http://dx.doi.org/10.1117/1.JBO.17.8.085003.1].

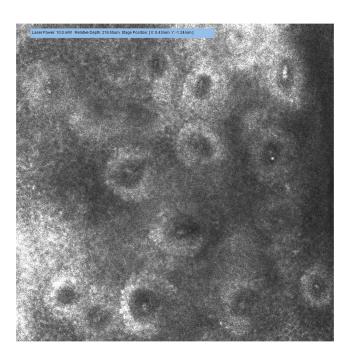


Video 2 Papillary dermis vascular circulation 10 min after capsaicin application (MPEG, 5.37 MB) [URI: http://dx.doi.org/10.1117/1.JBO .17.8.085003.2].

with the vascular response having a distinct aspect in various skin layers.³⁷ This is why *in vivo* RCM, considered an excellent method of evaluation, particularly for capillary vessels situated at the dermoepidermal junction,³⁵ may constitute a reference technique for future investigation of specific mechanisms through which capsaicin acts in the superior part of cutaneous microvascularization.



Video 3 Papillary dermis vascular circulation 25 min after capsaicin application (MPEG, 5.64 MB) [URI: http://dx.doi.org/10.1117/1.JBO .17.8.085003.3].



Video 4 Papillary dermis vascular circulation 40 min after capsaicin application (MPEG, 5.23 MB) [URI: http://dx.doi.org/10.1117/1.JBO .17.8.085003.4].

Administration of capsaicin, dissolved in immersion oil, allows uniform distribution of the active substance on the investigated skin area and strict control of quantity per unit of skin. Moreover, it facilitates image acquisition from the same skin area at each experimental stage. These characteristics may allow the development of a test with clinical applicability, as previous studies revealed that evaluation of neurogenic vascular reaction may constitute a valid test for small-diameter cutaneous nerve fiber functionality.³⁸ The test could be useful for early

diagnosis and quantification of therapeutic response in various types of neuropathies affecting thin cutaneous nerve fibers, such as diabetic neuropathy.

Investigation of micromorphological aspects of cutaneous neurogenic inflammation using *in vivo* RCM offers useful information for practitioners and researchers. It could be a potential diagnostic tool for cutaneous inflammatory disorders and could enable identification of new mechanisms by which the nervous system can interfere with physiopathological processes in skin inflammation.

Acknowledgments

Research funded by grant PN-II-RU-TE-2011-3-0249 from the National University Research Council, Romania. This study is part of the doctoral thesis of Constantin Caruntu that expresses his gratitude for Professor Leon Zagrean from "Carol Davila" Medicine and Pharmacy University, Bucharest for his guidance and support. Authors wish to thank Dr. Manuela Stanculescu from the Research Institute for Quality of Life, Romanian Academy for statistical analysis and Mihaela Adriana Ghita for her excellent assistance during the course of this work. Special thanks are given to Dr. Monica Neagu from "Victor Babes" Institute, Bucharest for revising the manuscript.

References

- S. P. Raychaudhuri and S. K. Raychaudhuri, "Role of NGF and neurogenic inflammation in the pathogenesis of psoriasis," *Prog. Brain Res.* 146, 433–437 (2004).
- W. Glinski et al., "Neuropeptides in psoriasis: possible role of betaendorphin in the pathomechanism of the disease," *Int. J. Dermatol.* 33(5), 356–360 (1994).
- T. Scholzen et al., "Neuropeptides in the skin: interactions between the neuroendocrine and the skin immune systems," *Exp. Dermatol.* 7(2–3), 81–96 (1998).
- C. Liezmann, B. Klapp, and E. M. Peters, "Stress, atopy and allergy: a re-evaluation from a psychoneuroimmunologic persepective," *Dermatoendocrinol.* 3(1), 37–40 (2011).
- 5. M. Ohshima et al., "Development of mechanisms associated with neurogenic-mediated skin inflammation during the growth of rats," *Pediatr. Res.* **67**(4), 363–368 (2010).
- A. Järvikallio, I. T. Harvima, and A. Naukkarinen, "Mast cells, nerves and neuropeptides in atopic dermatitis and nummular eczema," *Arch. Dermatol. Res.* 295(1), 2–7 (2003).
- F. Guarneri and H. Marini, "An unusual case of perioral dermatitis: possible pathogenic role of neurogenic inflammation," *J. Eur. Acad. Dermatol. Venereol.* 21(3), 410–412 (2007).
- S. Akaishi, R. Ogawa, and H. Hyakusoku, "Keloid and hypertrophic scar: neurogenic inflammation hypotheses," *Med. Hypotheses.* 71(1), 32–38 (2008).
- M. Steinhoff et al., "Clinical, cellular, and molecular aspects in the pathophysiology of rosacea," *J. Investig. Dermatol. Symp. Proc.* 15(1), 2–11 (2011).
- B. Zegarska, A. Lelińska, and T. Tyrakowski, "Clinical and experimental aspects of cutaneous neurogenic inflammation," *Pharmacol. Rep.* 58(1), 13–21 (2006).
- 11. C. Căruntu et al., "The role of stress in skin diseases," *Medicina Intern*ă *Internal Medecine*. **8**(3), 73–84 (2011).
- 12. C. Căruntu et al., "The role of stress in the multifactorial etiopathogenesis of acne," *Ro. Med. J.* **58**(2), 98–101 (2011).
- S. Lutgendorf et al., "Effects of relaxation and stress on the capsaicininduced local inflammatory response," *Psychosom. Med.* 62(4), 524–534 (2000).
- M. J. Caterina et al., "The capsaicin receptor: a heat-activated ion channel in the pain pathway," *Nature* 389(6653), 816–824 (1997).
- P. Holzer, "Local effector functions of capsaicin-sensitive sensory nerve endings: involvement of tachykinins, calcitonin gene-related peptide and other neuropeptides," *Neuroscience* 24(3), 739–768 (1988).

- J. D. Richardson and M. R. Vasko, "Cellular mechanisms of neurogenic inflammation," *J. Pharmacol. Exp. Ther.* 302(3), 839–845 (2002).
- D. Roosterman et al., "Neuronal control of skin function: the skin as a neuroimmunoendocrine organ," *Physiol. Rev.* 86(4), 1309–1379 (2006)
- B. Veronesi and M. Oortgiesen, "The TRPV1 receptor: target of toxicants and therapeutics," *Toxicol Sci.* 89(1), 1–3 (2006).
- A. Szallasi and P. M. Blumberg, "Vanilloid (Capsaicin) receptors and mechanisms," *Pharmacol Rev.* 51(2), 159–212 (1999).
- S. R. Sinclair et al., "Inhibition of capsaicin-induced increase in dermal blood flow by the oral CGRP receptor antagonist, telcagepant (MK-0974)," Br. J. Clin. Pharmacol. 69(1), 15–22 (2010).
- H. Wang et al., "Ethnic differences in pain, itch and thermal detection in response to topical capsaicin: African Americans display a notably limited hyperalgesia and neurogenic inflammation," *Br. J. Dermatol.* 162(5), 1023–1029 (2010).
- A. J. Terkelsen, F. W. Bach, and T. S. Jensen, "Experimental forearm immobilization in humans reduces capsaicin-induced pain and flare," *Brain Res.* 1263, 43–49 (2009).
- S. A. Boudreau et al., "Vascular and psychophysical effects of topical capsaicin application to orofacial tissues," *J. Orofac. Pain.* 23(3), 253–264 (2009).
- D. P. Stephens et al., "The influence of topical capsaicin on the local thermal control of skin blood flow in humans," Am. J. Physiol. Regul. Integr. Comp. Physiol. 281(3), R894–R901 (2001).
- P. Mohammadian, O. K. Andersen, and L. Arendt-Nielsen, "Correlation between local vascular and sensory changes following tissue inflammation induced by repetitive application of topical capsaicin," *Brain Res.* 792(1), 1–9 (1998).
- W. Koppert et al., "The effects of intradermal fentanyl and ketamine on capsaicin-induced secondary hyperalgesia and flare reaction," *Anesth. Analg.* 89(6), 1521–1527 (1999).

- J. Serra, M. Campero, and J. Ochoa, "Flare and hyperalgesia after intradermal capsaicin injection in human skin," *J. Neurophysiol.* 80(6), 2801–2810 (1998).
- B.J. Van der Schueren et al., "Reproducibility of the capsaicin-induced dermal blood flow response as assessed by laser Doppler perfusion imaging," Br. J. Clin. Pharmacol. 64(5), 580–590 (2007).
- C. Căruntu et al., "Modern techniques for the assessment of local skin blood flow," (Abstr.) Acta Medica. Marisiensis. 57(Suppl. 2), 8 (2011).
- C. Longo et al., "Skin aging: in vivo microscopic assessment of epidermal and dermal changes by means of confocal microscopy," J. Am. Acad. Dermatol. in press (2011).
- 31. P. Calzavara-Pinton et al., "Reflectance confocal microscopy for *in vivo* skin imaging," *Photochem. Photobiol.* **84**(6), 1421–1430 (2008).
- 32. A. Diaconeasa et al., "The role of confocal microscopy in the dermatooncology practice," *J. Med. Life.* 4(1), 63–74 (2011).
- M. A. Altintas et al., "Insight in human skin microcirculation using in vivo reflectance-mode confocal laser scanning microscopy," J. Digit. Imag. 23(4), 475–481 (2010).
- M. A. Altintas et al., "In vivo reflectance-mode confocal microscopy provides insights in human skin microcirculation and histomorphology," Comput. Med. Imag. Graph. 33(7), 532–536 (2009).
- 35. J. Hegyi et al., "Confocal laser-scanning capillaroscopy: a novel approach to the analysis of skin capillaries *in vivo*," *Skin Res. Technol.* **15**(4), 476–481 (2009).
- T. Yamashita et al., "In vivo assessment of pigmentary and vascular compartments changes in UVA exposed skin by reflectance-mode confocal microscopy," Exp. Dermatol. 16(11), 905–911 (2007).
- C. Forster et al., "Neurogenic flare responses are heterogeneous in superficial and deep layers of human skin," *Neurosci. Lett.* 185(1), 33–36 (1995).
- 38. A. Caselli et al., "Validation of the nerve axon reflex for the assessment of small nerve fibre dysfunction," *J. Neurol. Neurosurg. Psychiatry.* 77(8), 927–932 (2006).