Optical imaging in microfluidic bioreactors enables oxygen monitoring for continuous cell culture

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Abstract. For the first time, a fluorescence lifetime calibration method for an oxygen-sensitive dye ruthenium tris(2,2'-dipyridyl) dichloride hexahydrate (RTDP) is applied to image oxygen levels in poly(dimethyl siloxane) (PDMS) bioreactors containing living C2C12 mouse myoblasts. PDMS microsystems are broadly used in bioengineering applications due to their biocompatibility and ease of handling. For these systems, oxygen concentrations are of significance and are likely to play an important role in cell behavior and gene expression. Fluorescence lifetime imaging microscopy (FLIM) bases image contrast on fluorophore excited state lifetimes, which reflect local biochemical dynamics. Unique attributes of the widefield, time-domain FLIM system include tunable excitation (337.1 to 960 nm), large temporal dynamic range (≥600 ps), high spatial resolution (1.4 μm), calibrated detection (0 to 300±8 μM of oxygen), and rapid data acquisition and processing times (10 s). Oxygen levels decrease with increasing cell densities and are consistent with model outcomes obtained by simulating bioreactor oxygen diffusion and cell proliferation. In single bioreactor loops, FLIM detects spatial heterogeneity in oxygen levels with variations as high as 20%. The fluorescence lifetime-based imaging approach we describe avoids intensity-based artifacts (including photobleaching and concentration variations) and provides a technique with high spatial discrimination for oxygen monitoring in continuous cell culture systems.

Keywords: fluorescence microscopy; lifetime sensing; microfluidics; oxygen; molecular imaging; cell culture.

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Microfluidic devices have promising applications in cell-based assays and microscale tissue engineering, where spatiotemporal conditions are readily manipulated. Recently, poly(dimethyl siloxane)- (PDMS)-based microfluidic systems have been developed as biocompatible and rapidly prototyped systems for microscale-cell culture. For example, cells could be seeded and cultured successfully under continually perfused conditions to achieve an extracellular fluid-to-cell (volume) ratio close to the physiological value of 0.5. This small ratio facilitates heterogeneous chemical distribution, which may be critical in specifying cell fate in developing tissues. It is hence of great interest to quantitatively and with minimal perturbation characterize components (e.g., mitogens, nutrients, oxygen) in microfluidic bioreactors that influence cellular responses.

Oxygen in cell cultures influences cell signaling, growth, differentiation, and death. PDMS bioreactors are popular due to their high diffusivity of oxygen, which has been repeatedly demonstrated. It has been observed, however, that the diffusivity of PDMS can vary, depending on protein adsorption (e.g., when cells are cultured) or surface modification (e.g., plasma oxidation for bioreactors). It is hypothesized that this variability in PDMS permeability, along with cellular uptake and culture media perfusion, can affect spatial variations in oxygen within PDMS bioreactors.

Optical measurements of oxygen sensitive agents have advantages over more traditional, electrode-based approaches that make them uniquely applicable for bioreactor systems: they are well suited for small volumes, are relatively nonperturbing, and do not consume oxygen during the measurement. For long-term cell culture, optical sensing enables time-lapse studies (hours or days) without disturbing the setup, as well as imaging spatial oxygen distributions, which is useful for long-term cell culture.

Fluorescence intensity studies based on the oxygen sensitivity of ruthenium complexes embedded in matrix have been performed; these studies were intensity based, did not employ imaging, and did not report local oxygen concentration variations. Intensity-based fluorescence measurements are sensitive to instrumental variations (changes in excitation intensity or optical loss) and are affected by fluorophore concentration and photobleaching. Fluorescence lifetime, however, is an intrinsic property of the fluorophore’s excited electronic state and is insensitive to intensity artifacts. FLIM in microfluidic structures was reported for studying solvent mixing by monitoring viscosity in channels without cells. In this work, calibration of the oxygen sensitivity of RTDP fluorescence lifetime on a unique widefield, time-domain fluorescence lifetime imaging microscopy (FLIM) system was applied for quantitative oxygen estimation. To our knowledge, this is the first demonstration of using a lifetime imaging modality for extracellular oxygen monitoring in PDMS bioreactors containing living cells.

The FLIM system (Fig. 1) employs a tunable, pulsed (800-ps FWHM) excitation source operating in single-shot mode that covered the UV-vis-NIR spectrum (337 to 960 nm) and an intensified, gated (≥200 ps gate width) CCD camera (Picostar HR, LaVision) to record images, with an intensity reproducibility within ±2%. The system’s optical sectioning capability (via structured illumination) was not required for this study. Lifetime maps had a lateral resolution of 1.4 μm, which was well suited for studying small microfluidic struc-
photolithography. The seed channel was injected with the bioreactor was perfused by peristaltic pumping and the media reservoir 1 to 2 h after cell seeding. Oxygen was flowed over the sample and a drop in oxygen was observed extracellular oxygen level in the bioreactor decreased in a cell-density-dependent manner by almost 35% after incubation for 2 h. Oxygen levels measured here are lower compared to conventional macroscopic cell culture systems due to higher cell densities and low media levels. Lower oxygen levels are still conducive to cell growth and ideal for microscopic analysis. FLIM detection revealed that oxygen levels were taken at different points along each loop, as illustrated in Fig. 3. As expected, the observed extracellular oxygen level in the bioreactor decreased in a cell-density-dependent manner by almost 35% after incubation for 2 h. Oxygen levels measured here are lower compared to conventional macroscopic cell culture systems due to higher cell densities and low media levels. Lower oxygen levels are still conducive to cell growth in vitro, as indicated by proliferation of cells in the bioreactor over a 12 to 14 h period (data not shown).

A PDMS cell culture device with six loops is illustrated in Fig. 4(a). The device was small scale, optically transparent, and ideal for microscopic analysis. FLIM images of oxygen levels were taken at different points along each loop, as illustrated in Fig. 4(b). FLIM detection revealed that oxygen levels differed by as much as 20% within a loop, and this trend differed by as much as 20% within a loop, and this trend was consistently observed across multiple devices. Differences in measurements made at different points within the same loop were statistically significant [analysis of variance (ANOVA) and Student’s t test, p < 0.001]. (Comparisons across loops are not logical since each loop can potentially harbor different cell densities, which, as shown earlier, can significantly affect oxygen levels.)
Fig. 3 (Color online only) Simulation (white squares) and experimental FLIM (red squares) results of oxygen levels versus cell densities in channels illustrated in Fig. 2. Oxygen levels were estimated by averaging pixel values in oxygen distribution images of the channel. The model simulations were carried out according to the equations described in Ref. 11, with the model parameters set at maximum oxygen uptake rate \( V_{\text{m}} = 2 \times 10^{-16} \text{ mol/ cell s}^{-1} \), oxygen level at half-saturation \( K_{\text{m}} = 0.0059 \text{ mol/m}^3 \), overall mass transfer coefficient \( k_{\text{d}} = 4.5 \times 10^{-7} \text{ m/s} \), and estimated velocity of gravity flow \( (u) = 0.003 \text{ m/s} \). Error bars for some experimental data were within the red squares.

In conclusion, FLIM of RTDP was applied for the first time to measure oxygen in microfluidic bioreactors containing living cells. The calibrated method of oxygen estimation was sensitive over the entire physiological range (0 to 300 \( \mu \)M of oxygen) with a resolution of \( \pm 8 \text{ \mu M} \). Measured extracellular oxygen levels correlated with cell densities in the channels and were verified with model simulations (Figs. 2 and 3). Initial results on a bioreactor with six loops (Fig. 4) revealed statistically significant variations in oxygen levels within each bioreactor loop. An implicit assumption in FLIM measurements was that oxygenation did not vary significantly in the axial dimension due to the small height of the channel cross section (30 \( \mu \)m), an aspect that has been explored in more detail in simulation scenarios.11 Future work will involve long-term oxygen monitoring under continually perfused conditions. Modeling results indicate that media recycling might be beneficial to retain growth factor(s) secreted by cells; such a scheme will potentially affect oxygen levels, which can be tracked almost in real time due to fast data acquisition (<10 s) by the FLIM approach. All these endeavors are ultimately aimed at controlling long-term cellular responses (such as differentiation) in PDMS microbioreactors. A molecular, fluorescence-lifetime-based approach to oxygen sensing in PDMS bioreactors, such as that described here, offers several advantages including replacing a macroscopic chemical electrode with a nondestructive (and minimally perturbing) optical imager, which could potentially be customized and scaled-down using optoelectronic technologies, making this approach more economical and portable.

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**References**